

SUPPL. FIGURE LEGENDS

Supplemental Fig. S1. Exercise TRAINing attenuates weight gain and helps maintain muscle weight throughout DETRAINing.

A) Endurance exercise TRAIN paradigm in which mice fed a control diet (CD) underwent voluntary wheel running (VWR) or remained sedentary for 4wks. **B)** Body composition of the TRAIN cohort indicating stacked fat (yellow) and lean (pink) mass and body weight (lines, BW) changes. **C)** Weekly BW changes over time of the TRAIN cohort. **D)** Matched changes in fat (left) and lean (right) mass from 0 to 4wks. Inset above indicates a two-way ANOVA (time*exercise) showing significant effects with indicated p -values (1 cohort, $n=10$ /group). **E)** Relative muscle weights 2d post exercise of the TRAIN cohort. **F)** Absolute muscle weights 2d post exercise of the TRAIN cohort (1 cohort, $n=3$ /group). **G)** Endurance exercise TRAIN-DETRAIN paradigm in which mice trained for 4wks then detrained for 4wks or remained sedentary for 8wks. **H)** Body composition of the DETRAIN cohort indicating stacked fat (yellow) and lean (pink) mass and BW (lines) changes. **I)** Weekly BW changes from the DETRAIN cohort over 8wks. **J)** Matched changes in fat (left) and lean (right) after 4wks of TRAINing and 4wks of DETRAINing. Inset above indicates a two-way ANOVA (time*exercise) showing significant effects with indicated p -values (1 cohort, $n=10$ /group). **K)** Relative muscle weights from the DETRAIN cohort after 4wks of DETRAINing. **L)** Absolute muscle weights from the DETRAIN cohort after 4wks of DETRAINing (1 cohort, $n=3$ /group). **M)** Relative (left) and absolute (right) muscle weights of the Levator Ani (LA) 2d post completion of each cohort (4 cohorts, $n=3-6$ /group). **N)** Relative (left) and absolute (right) muscle weights of the Levator Ani (LA) 1wk post completion of each cohort (4 cohorts, $n=6-7$ /group). Data are presented as means \pm SD. Within timepoint comparisons determined by Student t-test; $^{\$}p<0.1$, $^*p<0.05$, $^{**}p<0.01$, $^{***}p<0.001$, $^{****}p<0.0001$.

Supplemental Fig. S2. RETRAINing tends to shift fiber distribution towards larger fibers with little impact on protein accretion.

A) Muscle fiber size frequency distribution of all fibers following TRAINing (left), DETRAINing (middle), and RETRAINing (right) (3 independent cohorts, 1 whole cross section per animal, $n=5-7$ /group). **B)** Representative western blot and **C)** analysis of puromycin incorporation (TA) into newly synthesized proteins (3 independent cohorts, $n=3$ /group). **D)** qPCR expression of genes related to protein degradation. Data are presented as means \pm SD. Within timepoint comparisons determined by Student t-test; $^{\$}p<0.1$, $^*p<0.05$, $^{**}p<0.01$, $^{***}p<0.001$, $^{****}p<0.0001$.

Supplemental Fig. S3. Effect of endurance training, detraining, and retraining on skeletal muscle gene expression.

Gastrocnemius RNA-seq showing the top 10 up- (red) and downregulated (blue) genes (left) and the KEGG enrichment analysis (right) following **A)** 4wks of endurance TRAINing (1 cohort, $n=3$ /group), **B)** 8wk DETRAINing (1 cohort, $n=3$ /group), and **C)** 12wk endurance RETRAINing cycles (1 cohort, $n=3$ /group) in VWR/CD mice, compared to age-matched SED/CD. DEGs adjusted $p<0.2$. *Tibialis anterior* qPCR expression of **D)** *Pgc-1 α* and **E)** epigenetic modifiers *Dnmt1*, *Tet1*, *Tet2*, and *Tet3* (3 independent cohorts, $n=6$ /group). Data are presented as means \pm SD. Within timepoint comparisons determined by Student t-test; $^{\$}p<0.1$, $^*p<0.05$, $^{**}p<0.01$, $^{***}p<0.001$, $^{****}p<0.0001$.

Supplemental Fig. S4. Exercise TRAINing negates HFD weight gain while increasing muscle growth, even following DETRAINing.

A) Mice fed a HFD and either exposed to 4wks voluntary wheel running (VWR) or static, sedentary cages (TRAIN). **B)** Body composition of the TRAIN cohort indicating stacked fat (yellow) and lean (pink) mass and body weight (BW, lines) changes. **C)** Weekly BW changes over time of the TRAIN cohort. **D)** Matched changes in fat (left) and lean (right) mass from 0 to 4wks from the TRAIN cohort. Inset above indicates a two-way ANOVA (time*exercise) showing significant effects with indicated p -values (1 cohort, $n=10$ /group). **E)** Relative muscle weights 2d post exercise of the TRAIN cohort. **F)** Absolute muscle weights 2d post exercise of the TRAIN cohort (1 cohort, $n=3$ /group). **G)** Mice were fed a HFD and completed 8wk TRAIN-DETRAIN with VWR or remained sedentary for 8wks (DETRAIN). **H)** Body composition of the DETRAIN cohort over 8wks indicating stacked fat (yellow) and lean (pink) mass and BW (lines) changes. **I)** Weekly BW changes of the DETRAIN cohort. **J)** Matched changes in fat (left) and lean (right) after 4wks of TRAINing and 4wks of DETRAINing. Inset above indicates a two-way ANOVA (time*exercise) showing significant effects with indicated p -values (1 cohort, $n=10$ /group). **K)** Relative muscle weights from the DETRAIN cohort after 4wks of DETRAINing. **L)** Absolute muscle weights from the DETRAIN cohort after 4wks of DETRAINing (1 cohort, $n=3$ /group). **M)** Relative (left) and absolute (right) muscle weights of the Levator Ani (LA) 2d post completion of each cohort (3 cohorts, $n=3-6$ /group). **N)** Relative (left) and absolute (right) muscle weights of the Levator Ani (LA) 1wk post completion of each cohort (3 cohorts, $n=6-7$ /group). Data are presented as means \pm SD. Within timepoint comparisons determined by Student t-test; $\$p<0.1$, * $p<0.05$, ** $p<0.01$, *** $p<0.001$, **** $p<0.0001$.

Supplemental Fig. S5. HFD feeding negates any increases in fiber size distribution, while RETRAINing may impact new protein accretion.

A) Muscle fiber size frequency distribution of all fibers following TRAINing (left), DETRAINing (middle), and RETRAINing (right) (3 independent cohorts, 1 whole cross section per animal, $n=6-7$ /group). **B)** Representative western blot and **C)** analysis of puromycin incorporation (TA) into newly synthesized proteins (3 independent cohorts, $n=3$ /group). **D)** qPCR expression of genes related to protein degradation. Data are presented as means \pm SD. Within timepoint comparisons determined by Student t-test; * $p<0.05$, ** $p<0.01$, *** $p<0.001$, $\$p<0.1$.

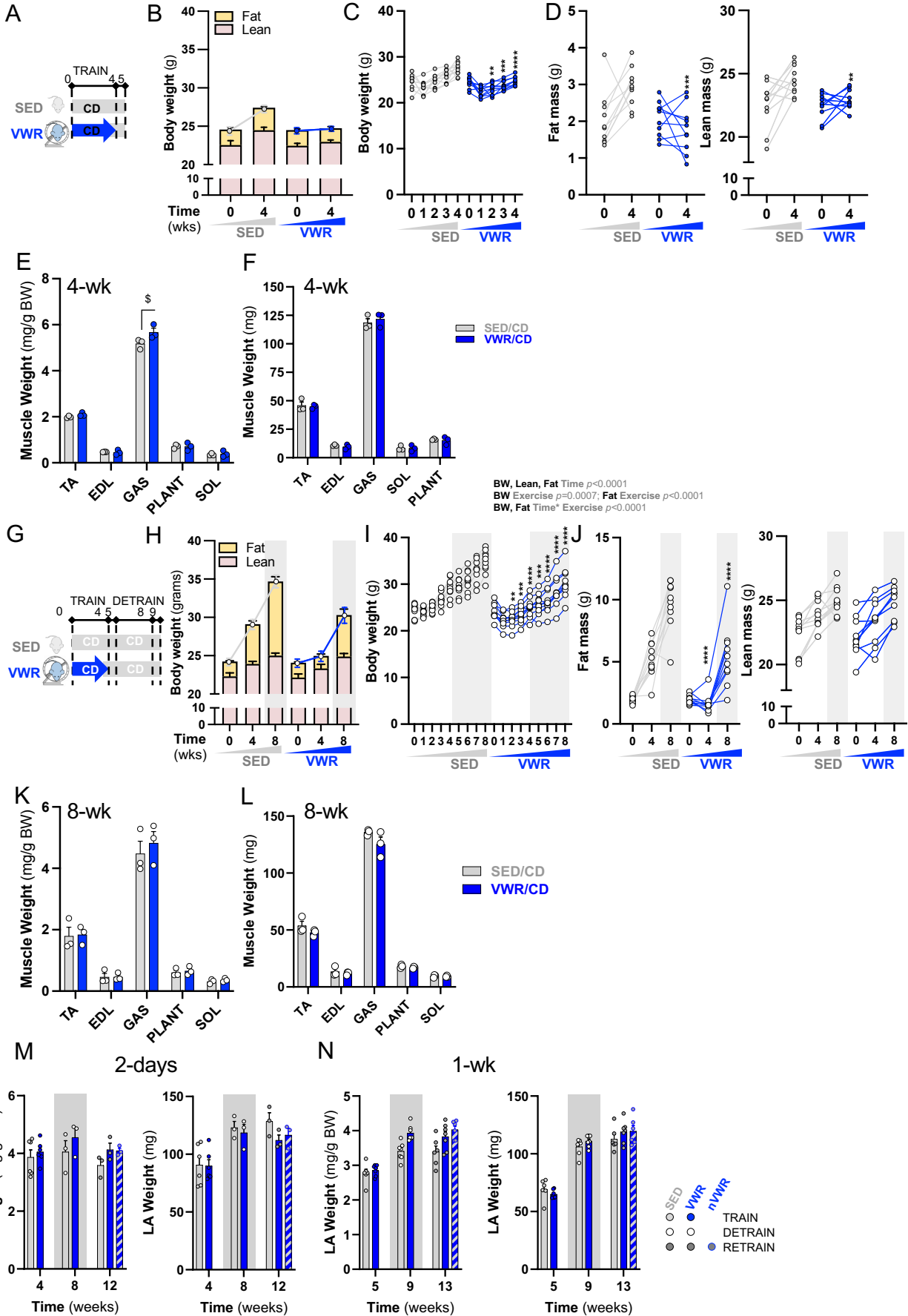
Supplemental Fig. S6. Impact of high-fat diet and exercise paradigms on skeletal muscle gene expression and pathway adaptations.

Gastrocnemius muscle RNA-seq showing the top 10 up- (red) and downregulated (blue) genes (left) and the KEGG enrichment analysis (right) comparing **A)** SED/HFD mice vs. SED/CD at 4wks, and **B)** 4wks of endurance TRAINing in VWR/HFD vs. SED/CD mice or, **C)** VWR/HFD vs. SED/HFD mice (1 cohort, $n=3$ /group). Muscle RNA-seq showing the top 10 up- (red) and downregulated (blue) genes (left) and the KEGG enrichment analysis (right) comparing **D)** SED/HFD mice vs. SED/CD at 8wks, and **E)** 8wks of DETRAINing in VWR/HFD vs. SED/CD mice or, **F)** VWR/HFD vs. SED/HFD mice (1 cohort, $n=3$ /group). Muscle RNA-seq showing the top 10 up- (red) and downregulated (blue) genes (left) and the KEGG enrichment analysis (right) comparing **D)** SED/HFD mice vs. SED/CD at 12wks, and **H)** 12wks of endurance RETRAINing in VWR/HFD vs. SED/CD mice or, **I)** VWR/HFD vs. SED/HFD mice (1 cohort, $n=3$ /group). DEGs adjusted $p<0.2$. *Tibialis anterior* qPCR expression of **J)** *Pgc-1 α* and **K)** epigenetic modifiers *Dnmt1*, *Tet1*, *Tet2*, and *Tet3* (3 independent cohorts, $n=6$ /group). Data are presented as means \pm SD. Within timepoint comparisons determined by Student t-test; $\$p<0.1$, * $p<0.05$, ** $p<0.01$, *** $p<0.001$, **** $p<0.0001$.

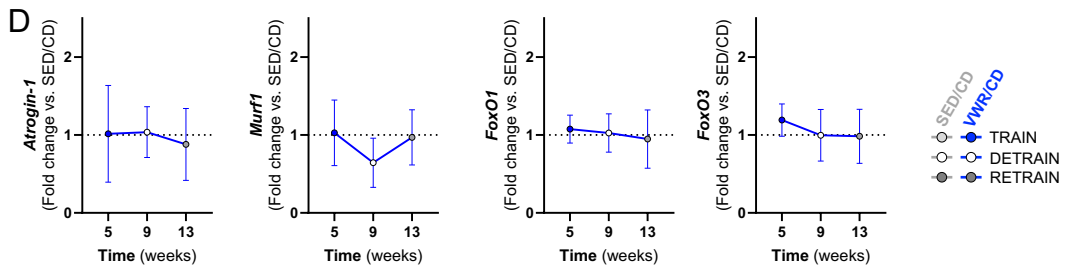
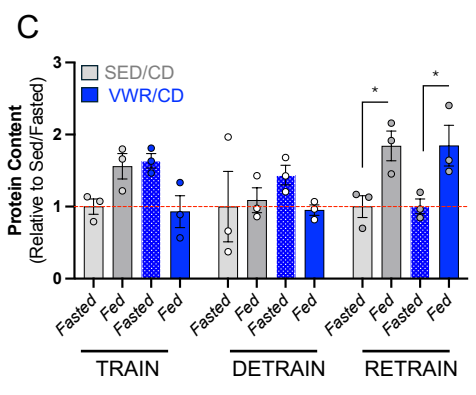
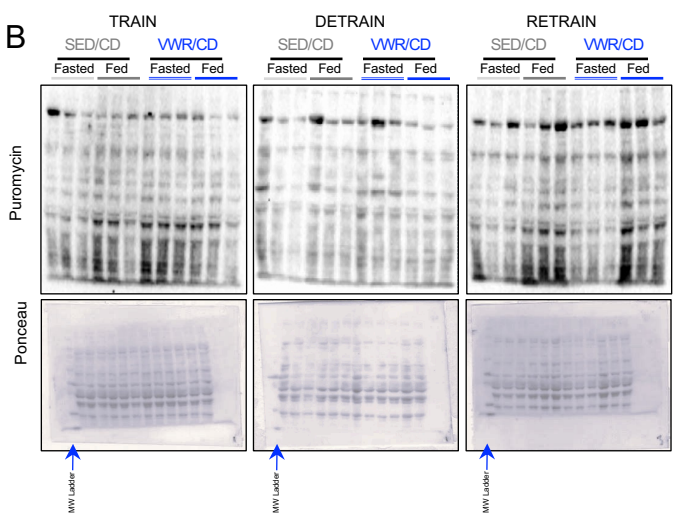
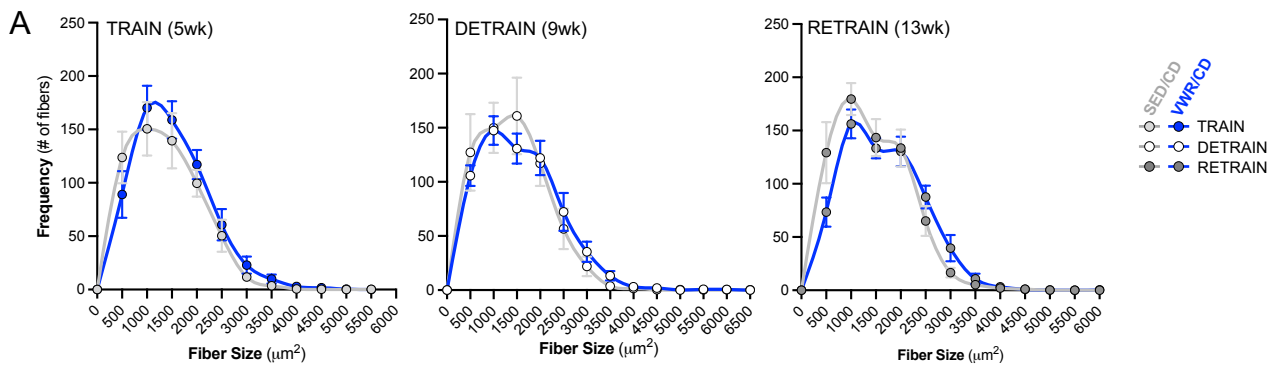
Supplemental Fig. S7. RETRAINing optimizes mitochondrial metabolism regardless of diet. Venn Diagram showing DEG overlap from VWR/CD, VWR/HFD, and SED/HFD following **A)** TRAINing, **B)** DETRAINing, and **C)** RETRAINing, compared to SED/CD. Upregulated (top; red, LogFC>1.0) and downregulated genes (bottom; blue, LogFC>1.0). DEGs adjusted $p < 0.2$. **D)** Sum of DEGs from at RETRAINing in both VWR/CD (vs. SED/CD) and VWR/HFD (vs. SED/HFD), and gene-network analysis for shared up- and downregulated genes.

Suppl. Fig. 1.

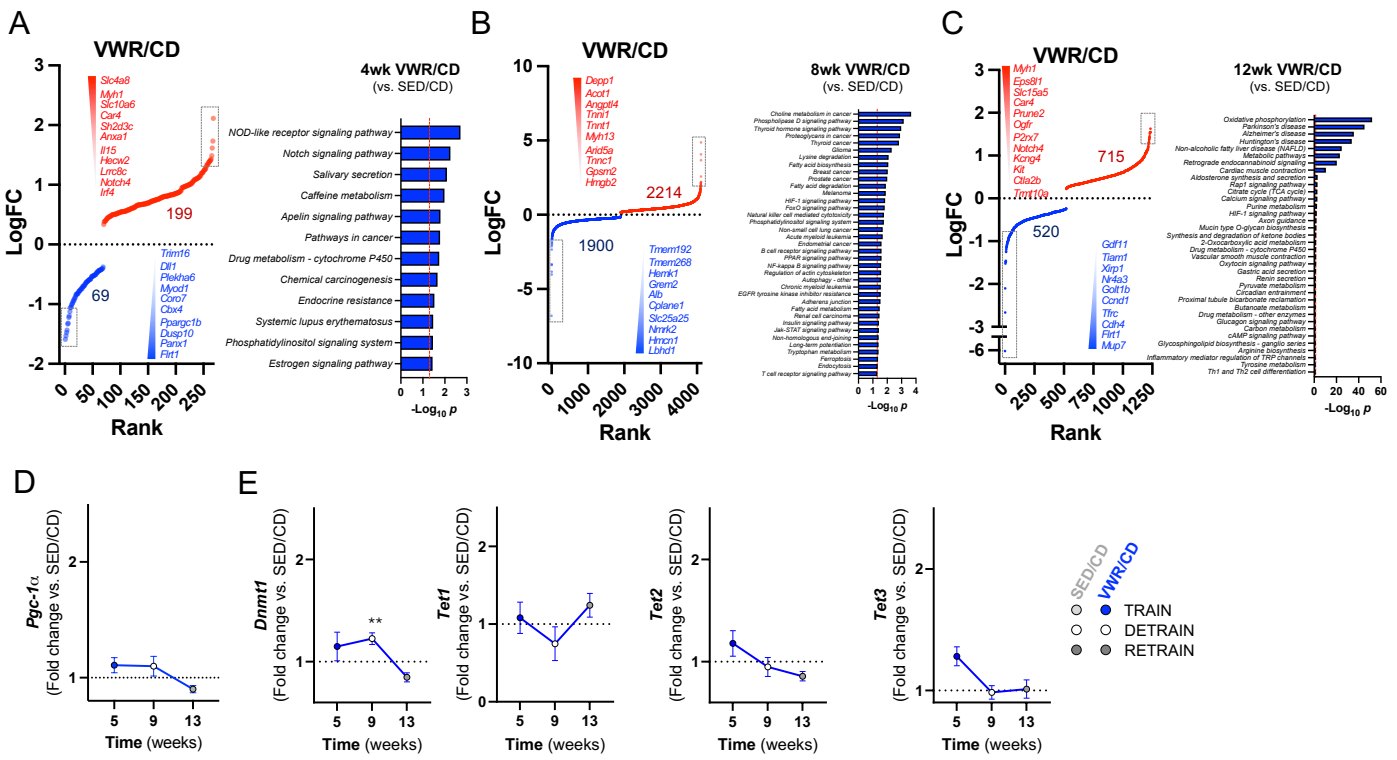
BW Time $p < 0.0001$; Lean Time $p = 0.0071$
 BW Exercise $p = 0.0011$; Fat Exercise $p = 0.0093$
 BW Time* Exercise $p = 0.0039$; Fat Time* Exercise $p = 0.0075$



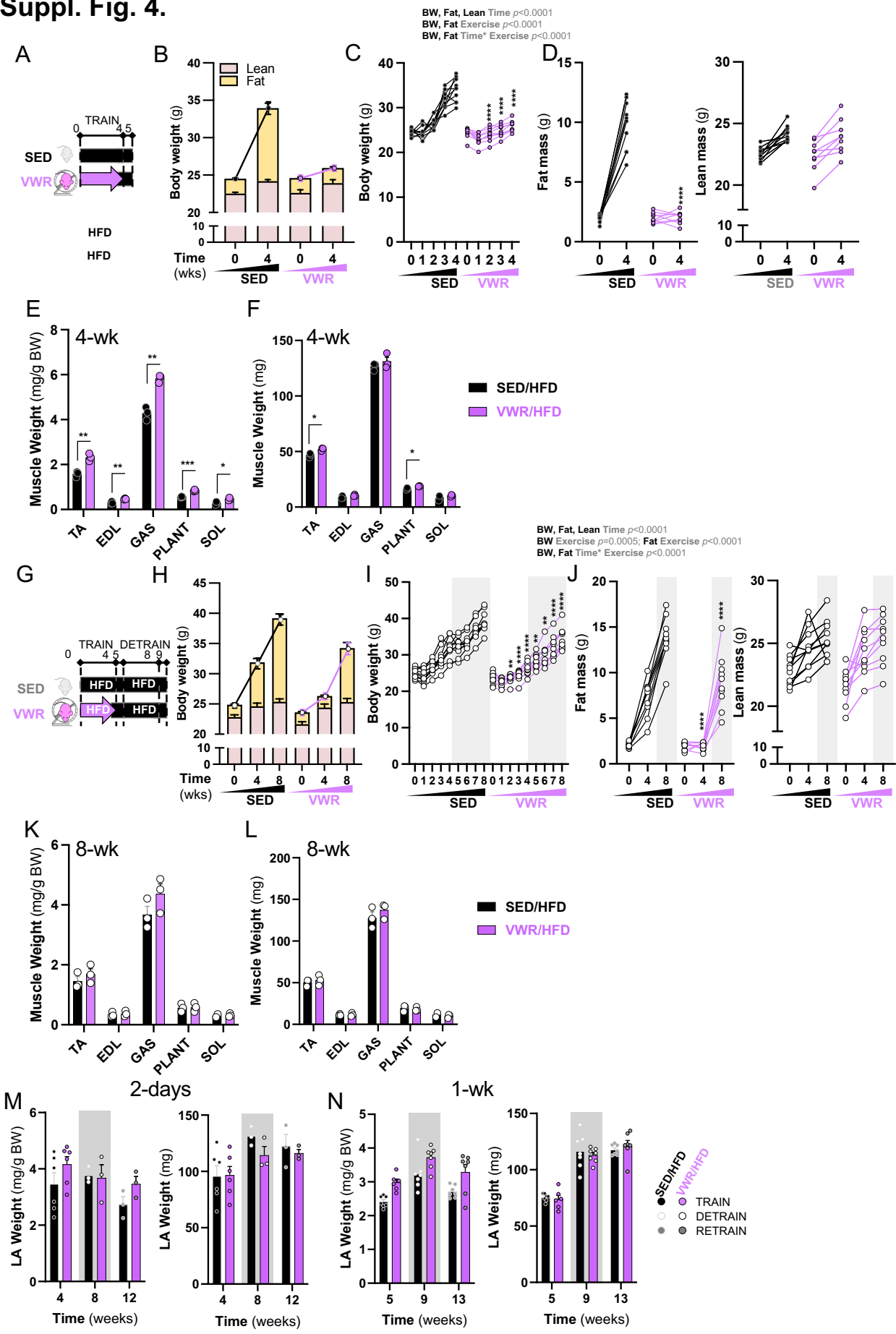
Suppl. Fig. 2.



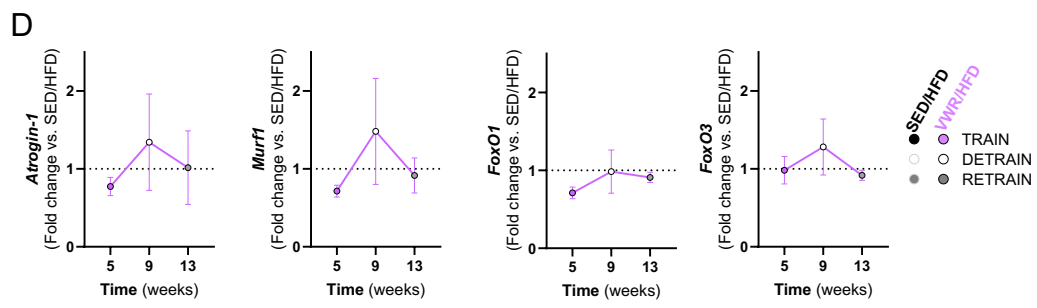
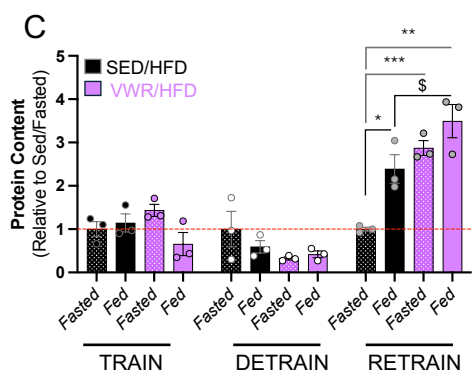
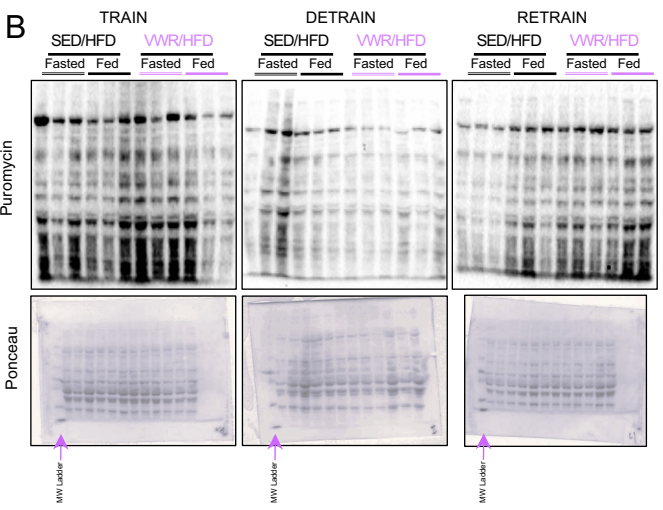
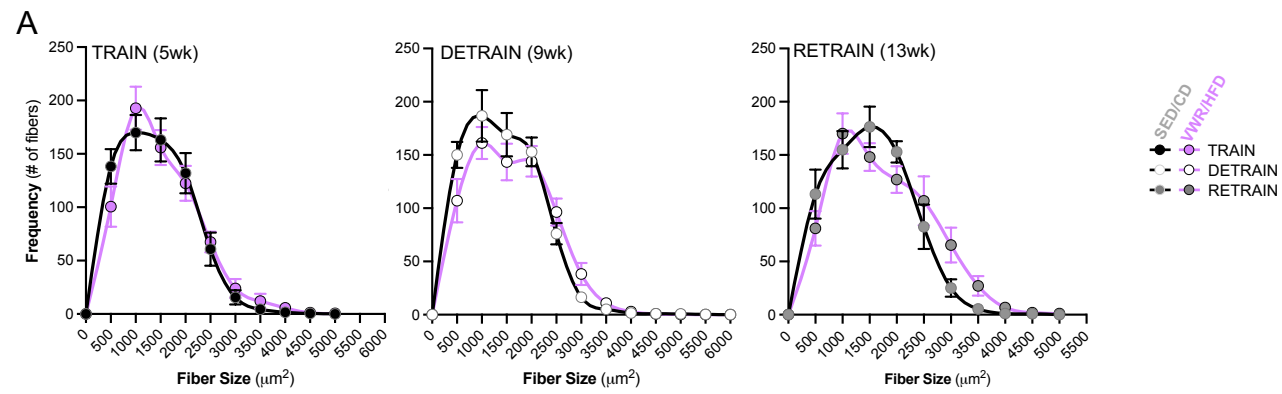
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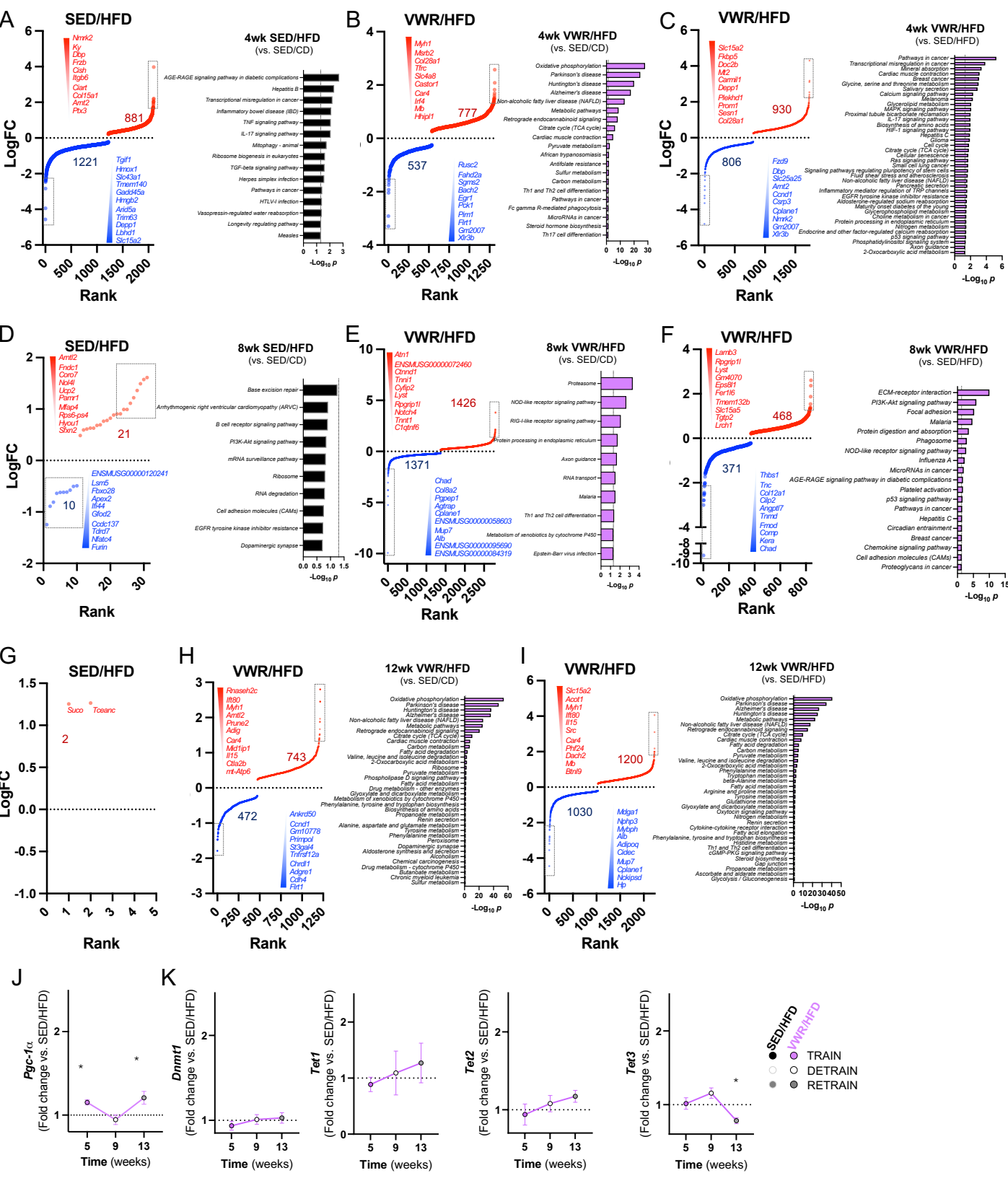
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Suppl. Fig. 5.



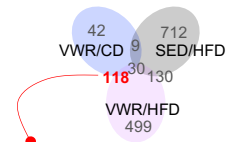
Suppl. Fig. 6.



Suppl. Fig. 7.

A

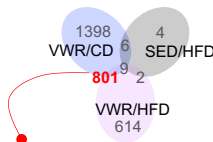
Upregulated (4wk)



LogFC > 1: *Slc4a8, Myh1, Slc10a6, Car4, Notch4, Irf4, Usp54, Cyp1b1, Plekhg1, Castor1, Rab27a, Csgalnact1, Arhgap25, Ifi205, Mef2l, Arhgap27, Adgrg1, Igfbp3, Lnx1, Rasgrp2, Scn2b, Dll4, Ccdc141, Xdh, Fmo2, Rsad2, ENSMUSG00000120992, Ehd3, Fbxl17, Kcna5, Kank3, Apobec3, Iqsec2, Btnl9.*

B

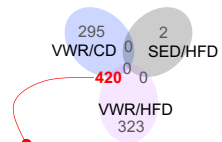
Upregulated (8wk)



LogFC > 1: *ENSMUSG00000072460, ENSMUSG00000107478, Tnni1, Tnnt1, Tnnc1, Myh7, Car4, Tpm3, ENSMUSG00000106037, Acsf2, P2rx7, Atp2a2, C1qtnf6, Cyfp2, Chn1, Ablim1, Btnl9, Lmod2, Sh2d3c, Cldn15, Myl2, Notch4, Rnf125, Lgi1, ENSMUSG00000099342, Abr, Med17, Pim3, N4bp3, Cyp1b1, Lipe, Cecr2, Aifm2, Impa2, Fmo2, Lnx1, Casc3, Zfp738, Shfl, Baz1a, Rnf144a, Kit, Slc39a14, Cd36, Rab15, Icam2, Liril4b, Sbd5, Spaar, Aldh9a1, Shank3, Cobll1, Wfs1, Fhl2, Rassf9, Tmem132b, Scube2.*

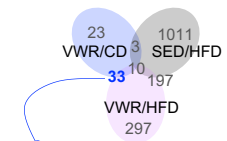
C

Upregulated (12wk)



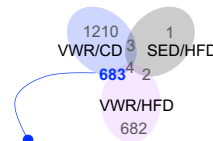
LogFC > 1: *Myh1, Slc15a5, Car4, Prune2, ENSMUSG00000102070, Ogr, ENSMUSG00000101249, Kcng4, Cta2b, Mid1p1, ENSMUSG00000089718, Cep85l, Dll4, Ckmt2, Ralgsd, Cacna1c, Cldn15, Adh1, Cxcr4, mt-Atp6, Kank3, Ii15, ENSMUSG00000097462, mt-Co3, Pde4c, ENSMUSG00000115970, Mtg1, Mb, Iqsec2, Tnfrsf10, Btnl9, Uqccl1, ENSMUSG00000100862, Apobec3, Cdh23, C1qtnf9, Xdh, Gstt2, mt-Nd3.*

Downregulated (4wk)



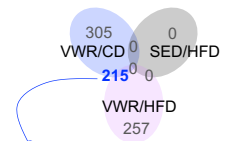
LogFC < -1: *Firt1, Myod1, Dll1, Trim16, Smad3, Cirbp, Rccd1, Plin2.*

Downregulated (8wk)



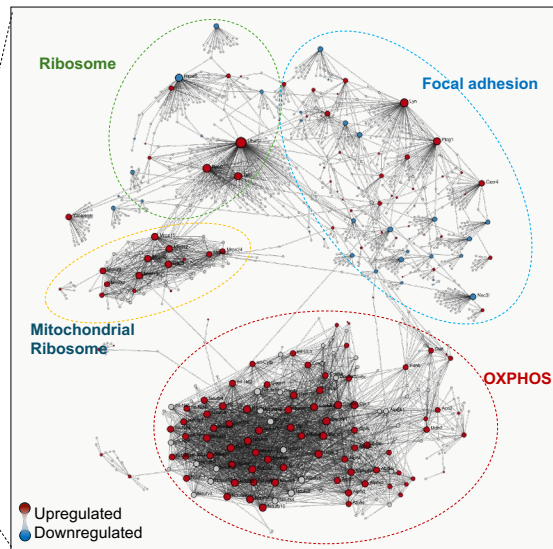
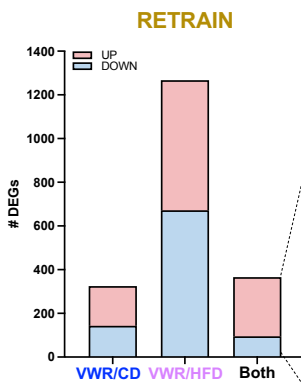
LogFC < -1: *ENSMUSG00000107244, ENSMUSG00000107585, ENSMUSG00000099552, Prkag3, Fam83d, Scn3b, ENSMUSG00000120512, Myod1, ENSMUSG00000121135, Gdf11, Nr4a1, Cntnap2, Usp37, Pgpep1, Ube2e1, Slc10a7, Mss51, ENSMUSG00000111556, Hemk1, Grem2, Alb, Cplane1, Slc25a25, ENSMUSG00000058603, ENSMUSG00000084319.*

Downregulated (12wk)



LogFC < -1: *ENSMUSG00000087579, Rasd2, Rbpj, ENSMUSG00000121486, Ankrd1, Xirp1, Nr4a3, ENSMUSG00000097125, Ccnd1, Cdh4, Firt1.*

D



TABLES

Supplemental Table S1. RT-qPCR primer sequences.

Gene	Forward sequence (5' to 3')	Reverse sequence (5' to 3')
<i>Pgc-1α</i>	GAATCAAGCCACTACAGACACCG	CATCCCTCTTGAGCCTTTTCGTG
<i>Dnmt1</i>	GGACAAGGAGAATGCCATGAAGC	TTACTCCGTCCAGTGCCACCAA
<i>Tet1</i>	CCCCAACTCCCACCAAACCC	AAACCCTTGTGGGACGGCTG
<i>Tet2</i>	ACCTGGCTACTGTCATTGCTCC	TGCAGTGACTCCTGAGAATGGC
<i>Tet3</i>	GGCCGATGCAGTAGTGGAG	TGAGCTCTGAGCCTGTCTTGA
<i>Atrogin-1</i>	CTTCTCGACTGCCATCCTGGAT	TCTTTTGGGCGATGCCACTCAG
<i>Murf1</i>	TACCAAGCCTGTGGTCATCCTG	ACGGAAACGACCTCCAGACATG
<i>FoxO1</i>	CTACGAGTGGATGGTGAAGAGC	CCAGTTCCTTCATTCTGCACTCG
<i>FoxO3</i>	CCTACTTCAAGGATAAGGGCGAC	GCCTTCATTCTGAACGCGCATG